# DEVELOPMENT OF ITERATIVE METHODS FOR THE EXACT DETERMINATION OF ACID DISSOCIATION CONSTANTS AND SPECIFIC REACTION RATES IN VARIOUS IONIC OR ELECTROCHEMICAL SYSTEMS. A REVIEW 

Georgios Papanastasiou ${ }^{1, *}$ and Georgios Kokkinidis ${ }^{2}$<br>Laboratory of Physical Chemistry, Department of Chemistry, Aristotle University of Thessaloniki, 54124 Thessaloniki, Greece; e-mail: ${ }^{1}$ papanast@chem.auth.gr, ${ }^{2}$ kokkinid@chem.auth.gr

Received February 25, 2003
Accepted May 24, 2003

This article is dedicated to Professor Sergio Roffia on the occasion of his retirement.


#### Abstract

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Theory and iterative methods for analysis of potentiometric titration data are presented in this paper. The methods concern titrations of (i) weak monoprotic acids, (ii) mixtures of weak acids with their conjugate bases (buffer solutions), (iii) mixtures of two monoprotic acids, (iv) solutions of polyprotic (di- and triprotic) acids with overlapping ionisation steps,
and (v) mixtures of two diprotic acids. Using equations derived without approximations and data exclusively resulting from the buffer region of a titration curve, it is possible to extract the accurate values of the concentration(s) and thermodynamic dissociation constant(s) of the titrated acid(s), even when the titration is stopped well before the end-point of the titration. The main principles of the iterative methods can also be applied to the evaluation of chronoamperometric curves for various electrochemical mechanisms (ECECE, ECE, irreversible, quasi-reversible, and CE). Thus, it is possible to extract the kinetic parameters $\mathrm{k}_{1}$ and $\mathrm{k}_{2}$ of an ECECE mechanism using exclusively chronoamperometric data. The analysis of the chronoamperometric curves for the other mechanisms (ECE, irreversible, quasi-reversible, and CE) permits the extraction of the corresponding kinetic parameters even when the values of the diffusion coefficient $D$ and the effective area $A$ of the electrode are not known. A review with 52 references.
Keywords: Acidity; Dissociation constants; Kinetics; Thermodynamics; Rate constants; Equilibria; Potentiometry; Chronoamperometry; Electrochemistry.

## 1. INTRODUCTION

Thermodynamic constants of acid-base equilibria ( $\mathrm{K}_{\text {diss }}$ ) and reaction rate constants (k) are very useful parameters for various chemical, electrochemical and technological applications. Some examples are given below.

It is known that catalytic methods can be used in analytical chemistry for determination of heavy metals. In those methods, weak acids, mainly polyprotic, are used. These acids, acting both as buffer and complexing agents, may enhance the catalytic action of the metal to be determined ${ }^{1-6}$, while, at the same time, they suppress the catalytic action caused by other metal ions present in solution. In order to assess the masking effect caused by an acid, it is necessary to know the stability constants of various complexes between the acid and metal ions. However, the determination of stability of such complexes demands accurate values of the dissociation constants of the acid used as complexing agent.

The production of compact and smooth metal deposits is an important task in electroplating processes. Metal deposits of uniform thickness and physical properties may be obtained if weak acids are used as additives in the plating solution. Moreover, these additives may prevent dendritic growth and, in some cases, can modify certain physical properties of the electrodeposits ${ }^{7-10}$. The influence of those additives has proved to be dependent on the extent of their dissociation ${ }^{11-13}$. Therefore, the dissociation constant $\mathrm{K}_{\text {diss }}$ appears to be a very useful parameter in explaining inhibition effects in the growth mechanism.

Finally, there is a large number of electrochemical reactions where both charge-transfer and chemical reactions are involved in the reaction mecha-
nism ${ }^{14}$. Typical electrode reactions with coupled homogeneous chemical reactions are the so-called CE, EC, ECE, and catalytic mechanisms ${ }^{14,15}$. Such systems are very common in organic electrochemistry. More complex mechanisms have also been reported. One example is the ECECE mechanism where three rapid charge-transfer steps are separated by two slow chemical reactions ${ }^{16-18}$. Depending on the sequence of steps involved and the values of the rate constants, the chemical steps may exert a dominant effect on the polarisation behaviour of the system.
The few examples reported above clearly show that thermodynamic constants of ionic equilibria or reaction rate constants are very useful parameters in quantitative analyses of various chemical and/or electrochemical systems. In this respect, techniques for accurate determination of such parameters have been developed during the last two decades in our laboratory. The basic principles of these methods are reviewed in the present article.

## 2. SIMULTANEOUS DETERMINATION OF EQUIVALENCE VOLUMES AND THERMODYNAMIC DISSOCIATION CONSTANTS OF WEAK ACIDS FROM POTENTIOMETRIC TITRATION DATA

The determination of the thermodynamic dissociation constants of weak acids can be realised by several methods. The most common procedure is the potentiometric titration. A variety of computational methods analysing potentiometric titration data were developed in the past few decades. Some of these methods are reported in excellent textbooks ${ }^{19-21}$.

The majority of the methods reported in literature are valid when: (i) the titrations are carried out under conditions of constant ionic strength and (ii) the accurate value of the analytical concentration of the titrated acid is known. The first condition is satisfied by addition of sufficient amounts of a neutral salt. However, the obtained pK values under such conditions are not free of any possible "salt effect". On the other hand, it has been argued in the literature that in media of high ionic strengths, even in the absence of a salt effect, the prediction of the activities of the various ionic species becomes inaccurate; the higher the ionic strength at which measurements are made, the more serious the difficulties in obtaining the dissociation constants become ${ }^{22}$. Concerning the analytical concentration of the titrated acid, this quantity is usually determined from the equivalence point of the titration curve. However, in some potentiometric titrations with a glass electrode in mixed water-organic solvent systems, the titrations curves of various weak acids do not show a pronounced vertical portion
corresponding to the end-point of neutralisation. On the other hand, the existence of multiple peaks in the immediate vicinity of the equivalence point of differential titration curves introduces an uncertainty in the choice of the exact value of the equivalence volume ${ }^{23}$. This behaviour could be attributed to two facts. First, the process of obtaining derivatives from experimental data usually results in considerably decreased accuracy ${ }^{24}$. Second, in the vicinity of the equivalence point, the response of the glass-calomel electrode set is slower than that observed in the buffer region of the same titration curve. Hence, in such solvent systems, it is very difficult to locate the end-point of the titration and to determine (or check) the accurate value of analytical concentration of the acid particular because of the contribution of hydrolysis of the salt formed. In addition, for some very weak acids, the equivalence volume does not coincide with the inflection point of the titration curves; the difference is the greater, the weaker is the acid ${ }^{19,25}$.

In an attempt to overcome these difficulties, new iterative techniques have been developed in our laboratory, permitting simultaneous determination of the thermodynamic constant and the analytical concentration of weak acids ${ }^{23,26}$. Thus, we have obtained theory and methods of analysis of potentiometric titration data for (i) weak monoprotic acids, (ii) mixtures of weak acids with their conjugate bases (buffer solutions), (iii) mixtures of two monoprotic acids, (iv) solutions of polyprotic (di- and triprotic) acids with overlapping ionisation steps, and (v) mixtures of two diprotic acids. These methods, using equations derived without approximations and data exclusively resulting from the buffer region of a titration curve, seem to be valid throughout the acid strength range. It is noted that all these methods can successfully be employed for titrations with absent sharp end-point, or for the determination of analytical concentrations and PK of a moderately weak acid in the presence of other very weak acids, the neutralisation of which is undesirable. The latter determination can easily be done by stopping the titration well before the end-point.

### 2.1. Potentiometric Titration of W eak M onoprotic Acids with Strong Bases

Consider a volume $\mathrm{V}_{0}$ of any weak monoprotic acid HA of initial concentration $\mathrm{C}_{\mathrm{o}}$, which is titrated with a strong base MOH of concentration $\mathrm{C}_{\mathrm{B}}$. Assuming that all singly charged ions have the same activity coefficient $\gamma_{1}$ and that the activity coefficients of uncharged species are unity, the thermodynamic acid constant of HA is given by Eq. (1).

$$
\begin{equation*}
\mathrm{K}_{\mathrm{a}}=\frac{\mathrm{a}_{\mathrm{H}}\left[\mathrm{~A}^{-}\right] \gamma_{1}}{[\mathrm{HA}]} \tag{1}
\end{equation*}
$$

At any point in the buffer region of the titration curve, the mass and charge balance are expressed by Eqs (2) and (3)

$$
\begin{equation*}
\mathrm{C}=[\mathrm{HA}]+\left[\mathrm{A}^{-}\right] \tag{2}
\end{equation*}
$$

$$
\begin{equation*}
\left[\mathrm{H}^{+}\right]+\left[\mathrm{M}^{+}\right]=\left[\mathrm{A}^{-}\right]+\left[\mathrm{OH}^{-}\right], \tag{3}
\end{equation*}
$$

where the concentrations $C$ and $\left[\mathrm{M}^{+}\right]$can be determined from the equations:

$$
\begin{gather*}
C=\frac{C_{B} V}{V_{o}+V}=b_{N} C_{B} V_{e}  \tag{4}\\
{\left[M^{+}\right]=\frac{C_{B} V_{e}}{V_{o}+V}=b_{N} C_{B} V,}
\end{gather*}
$$

where V is the volume of the added titrant, $\mathrm{V}_{\mathrm{e}}$ the equivalence volume and $b_{N}=1 /\left(V_{0}+V\right)$.

Combining these equations gives after rearrangement:

$$
\begin{equation*}
a_{H}\left(b_{N} C_{B} \vee \gamma_{1}+a_{H}-\frac{K_{w}}{a_{H}}\right)=K_{a}\left[b_{N} C_{B}\left(V_{e}-V\right)-\frac{a_{H}}{\gamma_{1}}+\frac{K_{w}}{a_{H} \gamma_{1}}\right] . \tag{5}
\end{equation*}
$$

$\mathrm{K}_{\mathrm{w}}$ is the autoprotolysis constant of the solvent used. For dilute solutions ( $<0.1 \mathrm{~m}$ ) the activity coefficient $\gamma_{1}$ can be calculated from the Debye-Hückel equation:

$$
\begin{equation*}
\log \gamma_{1}=-\frac{A z_{i}^{2} \sqrt{l}}{1+\mathrm{Ba}^{\circ} \sqrt{l}} \tag{6}
\end{equation*}
$$

where $A$ and $B$ are constants, the values of which depend on the physical properties of the medium ${ }^{27}, l$ is the ionic strength of the solution, $z_{i}$ the
charge number and $a^{\circ}$ the distance of closest approach of the ions. In principle, $a^{0}$ is a function of the solvent and electrolyte, but in the practice it is traditionally taken to be equal to $5 \AA^{20,28,29}$. The ionic strength, at any point of the titration curve, is given by Eq. (7) ${ }^{20,30}$ :

$$
\begin{equation*}
\mathrm{I}=\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]=\mathrm{b}_{\mathrm{N}} \mathrm{C}_{\mathrm{B}} \mathrm{~V}+\left[\mathrm{H}^{+}\right] . \tag{7}
\end{equation*}
$$

To determine $\gamma_{1}$, it is obviously necessary to know the value of $\left[\mathrm{H}^{+}\right]=$ $\mathrm{a}_{\mathrm{H}} / \gamma_{1}$ and vice versa. The values of $\left[\mathrm{H}^{+}\right]$and $\gamma_{1}$ can be obtained from the measured value of $\mathrm{a}_{\mathrm{H}}\left(\mathrm{pH}=-\log \mathrm{a}_{\mathrm{H}}\right)$ by successive approximations. Assuming first $\gamma_{1}=1$, so that $a_{H}=\left[\mathrm{H}^{+}\right]$, it is possible to calculate I from Eq. (7), and hence $\gamma_{1}$ from Eq. (6). This new value of $\gamma_{1}$ can be used to refine the values of $\left[\mathrm{H}^{+}\right]$, I and $\gamma_{1}$. All these operations are repeated until $\gamma_{1}$ converges (the criterion here being less than 0.00001 difference in I, between two subsequent cycles).

It is noted that Eq. (5), including the thermodynamic dissociation constant of the titrated acid, is also valid under conditions of varying ionic strength during the titration. Equation (5) has then the form

$$
\begin{equation*}
Y=K_{a} X . \tag{8}
\end{equation*}
$$

This equation predicts a linear relationship between $Y$ and $X$ with a slope equal to $\mathrm{K}_{\mathrm{a}}$. Alternatively, the experimental confirmation of such a correlation, using V and pH data, supports the assumption that the titrated acid is a monoprotic one. Then, the thermodynamic constant $\mathrm{K}_{\mathrm{a}}$ can be deduced, by linear regression, from the slope of the corresponding straight line.

However, there is no way in which, without prior knowledge of $\mathrm{V}_{\mathrm{e}}$, the Y vs $X$ data may be plotted to give a linear graph. However, this is possible with the iterative method proposed in this investigation. This procedure is analogous to previously proposed techniques for the determination of rate constants of SN2 reactions between ions and dipolar molecules in solvents where the ionic reactant associates to form ion pairs ${ }^{31-34}$.

### 2.1.1. Iterative $M$ ethod for the Determination of $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{K}_{\mathrm{a}}$

We assume first that the value of $\mathrm{V}_{\mathrm{e}}$ lies in an interval $(\mathrm{a}, \mathrm{b})$. Choosing arbitrarily from this interval a value for $\mathrm{V}_{\mathrm{e}}$, it is possible to trace, by means of
the experimental V and pH data, the curve $Y=f(X)$. This will approach a straight line to the extent where the chosen value of $\mathrm{V}_{\mathrm{e}}$ also approaches the exact value of the equivalence volume. So, the best linearity that could be obtained using the available experimental data, corresponds evidently to the best approximation to the exact value of the equivalence volume. So, by seeking the $\mathrm{V}_{\mathrm{e}}$ value within the interval $(\mathrm{a}, \mathrm{b})$, it is possible to trace N curves $Y=f(X), N$ being the number of estimations of $V_{e}$ values taken arbitrarily for these calculations. For each of these curves, the calculation of the squared correlation coefficient $R^{2}$ permits to compare the linearity of various $Y=f(X)$ plots and trace the curve $R^{2}=f\left(V_{e}\right)$. This curve is expected to present a pronounced maximum at a value of $\mathrm{V}_{\mathrm{e}}$ equal to $\mathrm{V}_{\mathrm{e}}{ }^{\text {max }}$ that can be considered as the best approximation to the exact value of the equivalence volume. The slope of the corresponding to $\mathrm{V}_{\mathrm{e}}{ }^{\text {max }}$ straight line is the best approximation to the exact value of $\mathrm{K}_{\mathrm{a}}$. It is noted that this iterative procedure is a general method that can be applied, as shown below, even to more complicated titrations (i.e., mixtures of two acids). However, in the titrations of weak monoprotic acids, $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{K}_{\mathrm{a}}$ can be more easily determined by the following graphical method.

### 2.1.2. Graphical Method for the Determination of $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{K}_{\mathrm{a}}$

Equation (5) can be rewritten as

$$
\begin{equation*}
y=V_{e}-\frac{1}{K_{a}} x, \tag{9}
\end{equation*}
$$

where

$$
\begin{equation*}
y=V+\frac{1}{\gamma_{1} C_{B} b_{N}}\left\{a_{H}-\frac{K_{w}}{a_{H}}\right\} \tag{10}
\end{equation*}
$$

and

$$
\begin{equation*}
x=a_{H}\left\{\gamma_{1} V+\frac{a_{H}}{C_{B} b_{N}}-\frac{K_{w}}{a_{H} C_{B} b_{N}}\right\} . \tag{11}
\end{equation*}
$$

In this treatment, it is possible to determine $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{K}_{\mathrm{a}}$ by linear regression from the coefficients of Eq. (9). It is noted that an analogous graphical procedure has been proposed by Johanson ${ }^{35}$. However, that approach focused rather on the determination of $\mathrm{V}_{\mathrm{e}}$, than of $\mathrm{K}_{\mathrm{a}}$. Indeed, in the Johanson method, the proposed equation, including the apparent dissociation con-
stant of the titrated acid, is valid under conditions where the ionic strength remains constant during the titration. This condition is normally fulfilled by addition of a neutral salt in sufficient amount. The difficulties arising in the determination of the thermodynamic $\mathrm{K}_{\mathrm{a}}$ in such media values were discussed previously.

### 2.2. Potentiometric Titration of a Solution Containing a Conjugate Acid-Base Pair

Consider a volume $\mathrm{V}_{0}$ of a solution containing a conjugate acid-base pair, where the initial analytical concentrations of the weak acid HA and its conjugate base are respectively equal to $\mathrm{C}_{0}$ and $\mathrm{C}_{\mathrm{b}}^{\circ}$. This mixture can be considered as being prepared by partial neutralisation of a solution of the acid HA, of concentration $\mathrm{C}_{0}^{\prime}$ and volume $\mathrm{V}_{\mathrm{o}}-\mathrm{Z}$, with Z ml of a solution of a strong base MOH of concentration $\mathrm{C}_{\mathrm{B}}$. At any point of the titration curve, mass and charge balance are expressed by the equations:

$$
\begin{gather*}
{\left[\mathrm{A}^{-}\right]+\left[\mathrm{OH}^{-}\right]=\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]}  \tag{12}\\
\mathrm{T}_{\mathrm{L}}=[\mathrm{HA}]+\left[\mathrm{A}^{-}\right], \tag{13}
\end{gather*}
$$

where

$$
\begin{gather*}
{\left[M^{+}\right]=\frac{C_{B} V}{V_{o}+V}+\frac{C_{b}^{\circ} V_{o}}{V_{o}+V}=b_{N} C_{B}(Z+V)}  \tag{14}\\
T_{L}=\frac{T_{L}{ }^{\circ} V_{o}}{V_{o}+V}+\frac{\left(Z+V_{e}\right) C_{B}}{V_{o}+V}=b_{N} C_{B}\left(Z+V_{e}\right) . \tag{15}
\end{gather*}
$$

Combining Eqs (1), (12)-(15), one obtains

$$
\begin{equation*}
\mathrm{y}=\mathrm{V}_{\mathrm{e}}-\frac{1}{\mathrm{~K}_{\mathrm{a}}} \mathrm{x}^{\prime} . \tag{16}
\end{equation*}
$$

In this equation, $y$ is again given by Eq. (10), while $x^{\prime}$ is determined from the following equation:

$$
\begin{equation*}
x^{\prime}=a_{H}\left\{\gamma_{1}(V+Z)+\frac{a_{H}}{C_{B} b_{N}}-\frac{K_{w}}{a_{H} C_{B} b_{N}}\right\} . \tag{17}
\end{equation*}
$$

At any point of the titration curve, the activity coefficient $\gamma_{1}$ can be calculated from Eq. (6), where the ionic strength is given by

$$
\begin{equation*}
\mathrm{I}=\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]=\mathrm{b}_{N} \mathrm{C}_{\mathrm{B}}(\mathrm{Z}+\mathrm{V})+\left[\mathrm{H}^{+}\right] . \tag{18}
\end{equation*}
$$

As previously, the values of $\left[\mathrm{H}^{+}\right]$and $\gamma_{1}$ may be obtained from the measured pH values by successive approximations.

Equation (16) predicts a linear relationship between $y$ and $x^{\prime}$ with a slope equal to $1 / K_{a}$. Alternatively, the experimental confirmation of such a correIation, using the V and pH data, supports the assumption that the titrated solution is a mixture of a monoprotic acid with its conjugate base. Then, $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{K}_{\mathrm{a}}$ can be deduced, by linear regression, from the coefficients of Eq. (16). This procedure can be realised by the presently proposed iterative method.

It is assumed first that the value of $Z$ lies in an interval $(a, b)$. Choosing arbitrarily from this interval a value for $Z$, it is possible to trace, using the experimental V and pH data, the curve $\mathrm{y}=\mathrm{f}\left(\mathrm{x}^{\prime}\right)$. This curve will approach a straight line to the extent where the chosen value of $Z$ also approaches the exact value of this parameter. Hence, the best linearity that could be obtained, using the available experimental data, evidently corresponds to the best approximation to the exact value of $Z$. Therefore, by seeking the $Z$ value within the interval ( $a, b$ ), it is possible to trace $N$ curves $y=f\left(x^{\prime}\right), N$ being the number of $Z$ values taken for these calculations. For each of these curves, the calculation of $R^{2}$ (square of the correlation coefficient) and $S_{y x}$ (standard error of the estimate) permits a comparison of the linearity of various $y=f\left(x^{\prime}\right)$ plots and trace the curves $R^{2}=f(Z)$ or $S_{y x}=f(Z)$. These curves must present respectively a maximum and minimum at a value of $Z$ equal to $Z_{m}$, which can be considered as the best approximation to the exact value of $Z$. The exact values of $V_{e}$ and $K_{a}$ can be deduced, respectively, from the $y$-intercept and the slope corresponding to $Z_{m}$ straight line.

### 2.3. Potentiometric Titration of Mixtures of Two W eak Monoprotic Acids with Strong Bases

The titration of mixtures of acids is a problem that was treated by numerous workers in the past. Two acids in a mixture with each other can simply be titrated if their pK values differ sufficiently. In such instances, the acids are neutralised serially in accord with their pK values. As a rule of thumb, a difference between the pK greater than 2 pK units is usually satisfactory to distinguish, by traditional titration, the component acids. For a smaller dif-
ference between the pK values, the acids are apparently neutralised simultaneously and the evaluation of the titration becomes increasingly difficult, yielding less reliable results. During the last decades, a wide variety of solutions to the problem were suggested ${ }^{36-42}$. All these methods require exact knowledge of the dissociation constants of the acids and most of them also require the sum of the concentrations of the acids. This situation has encouraged us to develop a new method permitting the simultaneous determination of the thermodynamic dissociation constants and the concentrations of two monoprotic acids in a mixture with each other even when the difference between their pK values is small. In this method, which requires only data resulting from the acid region of the titration curve, the sum of the concentrations need not be known.

Consider a volume $\mathrm{V}_{0}$ of a mixture of n weak monoprotic acids $\mathrm{HA}_{1}, \mathrm{HA}_{2}$, $\ldots, \mathrm{HA}_{n}$ of initial concentrations $\mathrm{C}_{1}^{0}, \mathrm{C}_{2}^{0}, \ldots, \mathrm{C}_{n}^{\circ}$ which is titrated with a solution of a strong base MOH of concentration $\mathrm{C}_{\mathrm{B}}$. Assuming again that all singly charged ions have the same activity coefficient $\gamma_{1}$ and that the activity coefficients of uncharged species are unity, the dissociation constant of the acid $\mathrm{HA}_{i}$ is expressed by Eq. (19):

$$
\begin{equation*}
\mathrm{K}_{\mathrm{i}}=\frac{\mathrm{a}_{\mathrm{H}}\left[\mathrm{~A}_{\mathrm{i}}^{-}\right] \gamma_{1}}{\left[\mathrm{HA}_{\mathrm{i}}\right]} . \tag{19}
\end{equation*}
$$

At any point of the acidic region of the titration curve, mass and charge balance are expressed by the following equations:
where

$$
\left.\begin{array}{c}
C_{\text {tot }}=\sum_{i=1}^{n}\left[\mathrm{HA}_{\mathrm{i}}\right]+\sum_{\mathrm{i}=1}^{n}\left[\mathrm{~A}_{\mathrm{i}}^{-}\right] \\
\sum_{\mathrm{i}=1}^{n}\left[\mathrm{~A}_{\mathrm{i}}^{-}\right]=\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]-\left[\mathrm{OH}^{-}\right]=\mathrm{F}
\end{array}\right\},
$$

$$
\begin{align*}
& C_{i}=\left[\mathrm{HA}_{\mathrm{i}}\right]+\left[\mathrm{A}_{\mathrm{i}}^{-}\right]=\left[\mathrm{HA}_{\mathrm{i}}\right]\left(1+\frac{\mathrm{K}_{\mathrm{i}}}{\mathrm{a}_{\mathrm{H}} \gamma_{1}}\right)=\mathrm{b}_{\mathrm{N}} \mathrm{C}_{\mathrm{i}}^{0} \mathrm{~V}_{0}  \tag{21c}\\
& \mathrm{~F}=\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]-\left[\mathrm{OH}^{-}\right]=\mathrm{b}_{\mathrm{N}} \mathrm{C}_{\mathrm{B}} \mathrm{~V}+\frac{\mathrm{a}_{\mathrm{H}}}{\gamma_{1}}-\frac{\mathrm{K}_{\mathrm{w}}}{\mathrm{a}_{\mathrm{H}} \gamma_{1}} \tag{21d}
\end{align*}
$$

Combination of Eqs (19)-(21) gives

$$
\begin{equation*}
C_{\text {tot }}-F=\sum_{i=1}^{n} \frac{C_{i}}{1+\frac{K_{i}}{a_{H} \gamma_{1}}} . \tag{22}
\end{equation*}
$$

For $n=2$, Eq. (22) yields

$$
\begin{equation*}
Y=A_{0}+A_{1} X_{1}+A_{2} X_{2}, \tag{23}
\end{equation*}
$$

where

$$
\left.\begin{array}{l}
A_{0}=K_{1} K_{2} \quad A_{1}=K_{1} \quad A_{2}=K_{2} \\
Y=\left(a_{H} \gamma_{1}\right)^{2} \frac{F}{L} \quad X_{1}=a_{H} \gamma_{1} \frac{b_{N} C_{1}^{o} V_{0}-F}{L} \quad X_{2}=a_{H} \gamma_{1} \frac{b_{N}\left(C_{\text {tot }}^{\circ}-C_{1}^{o}\right) V_{0}-F}{L}  \tag{24}\\
L=C_{\text {tot }}-F=b_{N} C_{B}\left(V_{e}-V\right)-\frac{a_{H}}{\gamma_{1}}+\frac{K_{w}}{a_{H} \gamma_{1}}
\end{array}\right\} .
$$

The activity coefficient $\gamma_{1}$ introduced in the above relationships can be determined, as previously, from Eq. (6) by successive approximations. Concerning the ionic strength $I$, a simple calculus gives

$$
\begin{equation*}
\mathrm{I}=\mathrm{b}_{\mathrm{N}} \mathrm{C}_{\mathrm{B}} \mathrm{~V}+\frac{\mathrm{a}_{\mathrm{H}}}{\gamma_{1}} . \tag{25}
\end{equation*}
$$

Equation (23) predicts that the relationship $Y=f\left(X_{1}, X_{2}\right)$ is linear. Alternatively, the experimental confirmation of such a correlation, using $V$ and pH data, supports the assumption that the titrated solution is a mixture of two monoprotic acids. Then, the thermodynamic acidity constants $K_{1}$ and
$\mathrm{K}_{2}$ may be obtained by a multiple linear regression method from the partial regression coefficients.

But, despite the fact that Eq. (23) offers a theoretical basis to correlate the results, there is no way in which, without prior knowledge of $\mathrm{V}_{\mathrm{e}}$ (or $\mathrm{C}_{\text {tot }}^{\circ}$ ) and $C_{1}^{\circ}$, the $X_{1}$ and $X_{2}$ data may be determined. This is possible with the iterative method proposed in this investigation.

This method is based on the particular property of Eq. (23) that, at the true value of the equivalence volume $\mathrm{V}_{\mathrm{e}}$ defined by Eq. (21b), this equation is linear, independent of the chosen value $\mathrm{C}_{1}^{0}$. Indeed, in the particular case where one uses the correct value $\mathrm{V}_{\mathrm{e}}$ and an erroneous value for $\mathrm{C}_{1}^{0}$ equal to $\hat{C}_{1}^{0}=C_{1}^{\circ}+\delta C_{1}^{\circ}$, the variables $X_{1}$ and $X_{2}$ are transformed into $\hat{X}_{1}$ and $\hat{X}_{2}$ defined by the following equations:

$$
\begin{gather*}
\hat{X}_{1}=a_{H} \gamma_{1} \frac{b_{N} \hat{C}_{1}^{\circ} V_{o}-F}{L}=X_{1}+\frac{a_{H} \gamma_{1} b_{N} V_{0}}{L} \delta \hat{C}_{1}^{\circ}  \tag{26}\\
\hat{X}_{2}=a_{H} \gamma_{1} \frac{b_{N}\left(C_{1}^{o}-\hat{C}_{1}^{o}\right) V_{0}-F}{L}=X_{2}-\frac{a_{H} \gamma_{1} b_{N} V_{0}}{L} \delta \hat{C}_{1}^{\circ} . \tag{27}
\end{gather*}
$$

Combination of Eqs (24), (26), and (27) yields

$$
\begin{equation*}
\frac{\mathrm{a}_{\mathrm{H}} \gamma_{1} \mathrm{~b}_{\mathrm{N}} \mathrm{~V}_{0}}{\mathrm{~L}}=\frac{\hat{X}_{1}-\hat{X}_{2}}{2\left(\mathrm{C}_{1}^{0}+\delta C_{1}^{0}\right)-C_{\text {tot }}^{0}} \tag{28}
\end{equation*}
$$

and combination of Eqs (23), (26)-(28) gives

$$
\begin{equation*}
Y=\hat{A}_{0}+\hat{A}_{1} \hat{X}_{1}+\hat{A}_{2} \hat{X}_{2} \tag{29}
\end{equation*}
$$

where

$$
\begin{gather*}
\hat{A}_{0}=A_{0}=\hat{K}_{1} \hat{K}_{2} \\
\hat{A}_{1}=K_{1}+\frac{\left(K_{2}-K_{1}\right) \delta C_{1}^{\circ}}{2\left(C_{1}^{\circ}+\delta C_{1}^{\circ}\right)-C_{\text {tot }}^{\circ}}  \tag{30}\\
\hat{A}_{2}=K_{2}-\frac{\left(K_{2}-K_{1}\right) \delta C_{1}^{\circ}}{2\left(C_{1}^{\circ}+\delta C_{1}^{\circ}\right)-C_{\text {tot }}^{\circ}}
\end{gather*}
$$

It follows from these equations that, using the true value of $\mathrm{V}_{\mathrm{e}}$ and any value for $C_{1}^{\circ}$, the relationship $Y=f\left(\hat{X}_{1}, \hat{X}_{2}\right)$ remains linear. However, the corresponding coefficients $\hat{\mathrm{A}}_{1}$ and $\hat{\mathrm{A}}_{2}$ are quite different from those of Eq. (23); the differences are greater, the greater is the value of $\delta C_{1}^{0}$. In the case when $\delta C_{1}^{\circ}=0$, so that the selected value for $\mathrm{C}_{1}^{0}$ is the correct one, both Eqs (29) and (23) are identical. This very important property may be used to determine the values of $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{C}_{1}^{0}$ as follows.

It is assumed first that $\mathrm{V}_{\mathrm{e}}$ is unknown and that its true value lies in an interval $(a, b)$. Let $\hat{C}_{1}^{\circ}$ be any trial value for $\mathrm{C}_{1}^{\circ}$ chosen arbitrarily from an interval ( $a^{\prime}, b^{\prime}$ ) of the possible values for this concentration. Starting from this value, and taking from the interval $(a, b)$ any value for $\mathrm{V}_{\mathrm{e}}$, it is possible to obtain, using the experimental $V$ and pH data, the dependence $Y=f\left(\hat{X}_{1}\right.$, $\left.\hat{X}_{2}\right)$. The latter will approach a linear relationship to the extent where the chosen $\mathrm{V}_{\mathrm{e}}$ also approaches the exact value of the equivalence volume. Thus, the best linearity which can be obtained, using the available experimental data, corresponds evidently to the best approximation to the exact value of the equivalence volume. Hence, by seeking within the interval $(a, b)$ the $\mathrm{V}_{\mathrm{e}}$ value, it is possible to obtain $N$ relationships $Y=f\left(\hat{X}_{1}, \hat{X}_{2}\right), N$ being the number of $V_{\mathrm{e}}$ values taken for these calculations. For each of these relationships, the calculation of the squared multiple correlation coefficient $R^{2}$ allows to compare the linearity of the various $Y=f\left(\hat{X}_{1}, \hat{X}_{2}\right)$ variations and to trace the curve $R^{2}=f\left(V_{e}\right)$. This is supposed to present a pronounced maximum at a value of $V_{e}$ equal to $V_{e}{ }^{\text {max }}$, which can be considered the best approximation to the exact value of the equivalence volume. At this value of $\mathrm{V}_{\mathrm{e}}$, the partial regression coefficients of the corresponding multiple regression equation will approach the coefficients of Eq. (23) to the extent where the chosen value $\hat{C}_{1}^{\circ}$ also approaches the exact value of $\mathrm{C}_{1}^{\circ}$. In a second step, starting from the value $\mathrm{V}_{\mathrm{e}}^{\text {max }}$, and seeking within the interval ( $\mathrm{a}^{\prime}, \mathrm{b}^{\prime}$ ) the $\hat{C}_{1}^{\circ}$ value, it is possible to obtain, as above, $N^{\prime}$ relationships $Y=f\left(\hat{X}_{1}\right.$, $\hat{X}_{2}$ ), $N^{\prime}$ being the number of $\hat{C}_{1}^{\circ}$ values taken for these calculations. For each of these relationships, it is possible to calculate the ratio

$$
\begin{equation*}
R_{c f}=\frac{\hat{A}_{0}}{\hat{A}_{1} \hat{A}_{2}} . \tag{31}
\end{equation*}
$$

This ratio may be equal to unity at a certain value of $\hat{\mathrm{C}}_{1}^{\circ}$ that can be considered as the best approximation to the exact value of $\mathrm{C}_{1}^{0}$. Evidently, $\mathrm{C}_{2}^{0}$ can be calculated from Eq. (21b), while $\mathrm{pK}_{1}$ and $\mathrm{pK}_{2}$ can be estimated from the coefficients of the corresponding multiple linear regression equation.

### 2.4. Potentiometric Titration of W eak Polyprotic Acids with Strong Bases

Consider a volume $\mathrm{V}_{0}$ of a solution of a molecular polyprotic acid $\mathrm{H}_{\mathrm{n}} \mathrm{A}$ with overlapping ionisation steps of initial concentration $\mathrm{C}_{0}$, which is titrated with a solution of strong base MOH of concentration $\mathrm{C}_{\mathrm{B}}$. The titration is assumed to be carried out in the acid range. In this range, the analysis of the titration curve leads to the following equation ${ }^{19}$

$$
\begin{equation*}
\hat{h}=\frac{\frac{K_{1}}{a_{H} \gamma_{1}}+\frac{2 K_{1} K_{2}}{a_{H}^{2} \gamma_{1}}+\ldots+\frac{n K_{1} K_{2} \ldots K_{n}}{a_{H}^{n} \gamma_{n}}}{1+\frac{K_{1}}{a_{H} \gamma_{1}}+\frac{K_{1} K_{2}}{a_{H}^{2} \gamma_{1}}+\ldots+\frac{K_{1} K_{2} \ldots K_{n}}{a_{H}^{n} \gamma_{n}}} \tag{32}
\end{equation*}
$$

with

$$
\begin{equation*}
\hat{h}=\frac{\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]-\left[\mathrm{OH}^{-}\right]}{\mathrm{C}} \tag{33}
\end{equation*}
$$

and

$$
\begin{equation*}
C=\frac{C_{0} V_{0}}{V_{0}+V} b_{N} C_{0} V_{0}=\frac{1}{n} b_{N} C_{B} V_{e}, \tag{34}
\end{equation*}
$$

where $\gamma_{i}$ denotes the activity coefficient of the species ( $i=0,1,2, \ldots, n$ ) and $\mathrm{K}_{\mathrm{i}}$ the dissociation constants of the various ionisation steps.

For diprotic acids, Eq. (32) gives

$$
\begin{equation*}
y=-K_{1}+K_{1} K_{2} x \tag{35}
\end{equation*}
$$

with

$$
\begin{equation*}
y=\frac{Z}{X} \quad \text { and } \quad x=\frac{Y}{X} \tag{36}
\end{equation*}
$$

and

$$
\left.\begin{array}{l}
X=\frac{\gamma_{2}\left[b_{N} C_{B}\left(V-\frac{V_{e}}{2}\right) \gamma_{1}+a_{H}-\frac{K_{w}}{a_{H}}\right] a_{H}}{\gamma_{1}} \\
Y=b_{N} C_{B}\left(V_{e}-V\right) \gamma_{1}+\frac{K_{w}}{a_{H}}-a_{H} \\
Z=\left(b_{N} C_{B} V \gamma_{1}+a_{H}-\frac{K_{w}}{a_{H}}\right) a_{H}^{2} \gamma_{2}
\end{array}\right\} .
$$

For dilute solutions $\left(I<10^{-1} \mathrm{M}\right)$, the activity coefficients $\gamma_{1}$ and $\gamma_{2}$ can be determined from Eq. (6), and the ionic strength I of the solution from the following relationship ${ }^{30}$ :

$$
\begin{equation*}
\mathrm{I}=\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]+\left[\mathrm{A}^{2-}\right]=\mathrm{b}_{N} \mathrm{C}_{\mathrm{B}} \mathrm{~V}+\frac{\mathrm{a}_{\mathrm{H}}}{\gamma_{1}}+\left(\frac{\mathrm{K}_{1} \mathrm{~K}_{2}}{a_{H}^{2} \gamma_{2}}\right) \frac{\mathrm{C}}{\mathrm{D}}, \tag{38}
\end{equation*}
$$

where

$$
\begin{equation*}
D=1+\frac{K_{1}}{a_{H} \gamma_{1}}+\frac{K_{1} K_{2}}{a_{H}^{2} \gamma_{2}} . \tag{39}
\end{equation*}
$$

The determination of $\mathrm{V}_{\mathrm{e}}, \mathrm{K}_{1}$, and $\mathrm{K}_{2}$ from Eq. (35), using V and pH data, is possible with the iterative method proposed here. It is assumed first that the value of $\mathrm{V}_{\mathrm{e}}$ lies in an interval ( $\mathrm{a}, \mathrm{b}$ ). Choosing arbitrarily from this interval a value for $\mathrm{V}_{\mathrm{e}}$, calculation of $\gamma_{1}$ and $\gamma_{2}$, at any point of the titration curve, requires of course the knowledge of the corresponding value of I. On the other hand, I is a function of $K_{1}, K_{2}, \gamma_{1}$, and $\gamma_{2}$. Thus, determination of $\gamma_{1}$ and $\gamma_{2}$ requires prior knowledge of $\gamma_{1}$ and $\gamma_{2}$. It appears that these calculations constitute a typical case of a vicious circle. However, $\gamma_{1}$ and $\gamma_{2}$ may be obtained by successive approximations. Assuming first that $\gamma_{1}$ and $\gamma_{2}$ equal 1 , it is possible to determine approximate values (apparent constants) of $K_{1}$, $\mathrm{K}_{2}$ from the coefficients of Eq. (35) using the experimental V and pH data.

Starting from these values and assuming again that $\gamma_{1}$ and $\gamma_{2}$ equal 1 , so that $\mathrm{a}_{\mathrm{H}}=\left[\mathrm{H}^{+}\right]$, it is possible to calculate $I$ at any chosen point of the titration curve, and hence $\gamma_{1}$ and $\gamma_{2}$ from Eq. (6). These new values $\gamma_{1}$ and $\gamma_{2}$ can
be used to refine values of I and to repeat this until it converges (the criterion being less than the $10^{-5}$ difference in I between two subsequent cycles). By repeating these approximations at each point of the titration curve, it is possible to trace the curve $y=f(x)$ and to determine by the least-squares method the values of $K_{1}$ and $K_{2}$. These new values can be used to refine the values of $K_{1}$ and $K_{2}$ and to repeat this until they converge (the criterion being less than the $10^{-5}$ difference in $\mathrm{pK}_{1}$ and $\mathrm{pK}_{2}$ values between two subsequent cycles). The final curve $y=f(x)$, obtained using the approximation, approaches a straight line to the extent the chosen value of $\mathrm{V}_{\mathrm{e}}$ also approaches the exact value of this parameter. Hence, the best linearity that could be obtained using the available experimental data evidently corresponds to the best approximation to the exact value of the equivalence volume. Therefore, by seeking within the interval $(a, b)$ the $V_{e}$ value, it is possible to trace N curves $\mathrm{y}=\mathrm{f}(\mathrm{x}), \mathrm{N}$ being the number of $\mathrm{V}_{\mathrm{e}}$ values taken for these calculations. For each of these curves, the calculation of $R_{2}$ and $S_{y x}$ permits the comparison of the adequacy of the fit of Eq. (35) to the experimental V and pH data. At the true value of $\mathrm{V}_{\mathrm{e}}$, these statistics may reach a maximum and a minimum value, respectively. Then, the exact values of $\mathrm{K}_{1}$ and $K_{2}$ can be deduced, respectively, from the coefficients of the corresponding straight line.

An analogous procedure can be used in order to analyse the titration curve for triprotic acids. Indeed, in this case Eq. (32) gives

$$
\begin{equation*}
Y=K_{1}+\left(K_{1} K_{2}\right) X_{1}+\left(K_{1} K_{2} K_{3}\right) X_{2} \tag{40}
\end{equation*}
$$

with

$$
\begin{equation*}
Y=\left(\frac{\hat{h}}{1-\hat{h}}\right) a_{\mathrm{H}} \gamma_{1}, \quad X_{1}=\left(\frac{2-\hat{h}}{1-\hat{h}}\right) \frac{\gamma_{1}}{a_{\mathrm{H}} \gamma_{2}}, \quad X_{2}=\left(\frac{3-\hat{h}}{1-\hat{h}}\right) \frac{\gamma_{1}}{a_{H}^{2} \gamma_{3}} . \tag{41}
\end{equation*}
$$

As previously, the activity coefficients $\gamma_{1}, \gamma_{2}$ and $\gamma_{3}$ can be determined from Eq. (6), where the ionic strength is given by the following formula ${ }^{43}$ :

$$
\begin{align*}
I & =\left[M^{+}\right]+\left[\mathrm{H}^{+}\right]+\left[\mathrm{HA}^{2-}\right]+3\left[\mathrm{~A}^{3-}\right]= \\
& =b_{N} C_{B} V+\frac{a_{H}}{\gamma_{1}}+\left(\frac{K_{1} K_{2}}{a_{H}^{2} \gamma_{2}}+\frac{K_{1} K_{2} K_{3}}{a_{H}^{3} \gamma_{3}}\right) \frac{C}{D}, \tag{42}
\end{align*}
$$

where

$$
\begin{equation*}
D=1+\frac{K_{1}}{a_{H} \gamma_{1}}+\frac{K_{1} K_{2}}{a_{H}^{2} \gamma_{2}}+\frac{K_{1} K_{2} K_{3}}{a_{H}^{3} \gamma_{3}} . \tag{43}
\end{equation*}
$$

The activity coefficients $\gamma_{1}, \gamma_{2}$, and $\gamma_{3}$ can be calculated by successive approximations, as described above.
The fact that Eq. (40) is linear, can be used for determination of $\mathrm{V}_{\mathrm{e}}, \mathrm{K}_{1}$, $K_{2}$, and $K_{3}$, using the iterative method described in detail for monoprotic and diprotic acids.

### 2.5. Potentiometric Titration of Mixtures of Two W eak Diprotic Acids with Strong Bases

The titration of mixtures of two weak diprotic acids was also treated by Purdie et al. elsewhere ${ }^{38}$. However, the Purdie method requires the knowledge of the sum of the concentrations of the acids, the pK values being known. In contrast, in the present method the sum of the concentrations need not be known.

Consider a volume $\mathrm{V}_{0}$ of a mixture of two weak diprotic acids $\mathrm{H}_{2} \mathrm{~A}_{1}$ and $\mathrm{H}_{2} \mathrm{~A}_{2}$ of initial analytical concentrations $\mathrm{C}_{1}^{\circ}$ and $\mathrm{C}_{2}^{\circ}$, which is titrated with a solution of a strong base MOH of concentration $\mathrm{C}_{\mathrm{B}}$. Assuming that all equally charged ions have the same activity coefficients, the dissociation constants $\mathrm{K}_{\mathrm{i} 1}, \mathrm{~K}_{\mathrm{i} 2}$ of the acid $\mathrm{H}_{2} \mathrm{~A}_{\mathrm{i}}$ (with $\mathrm{i}=1,2$ ) are given by

$$
\begin{equation*}
K_{i 1}=\frac{a_{H}\left[H A_{i}^{-}\right] \gamma_{1}}{\left[H_{2} A_{i}\right]}, \quad K_{i 2}=\frac{a_{H}\left[A_{i}^{2-}\right] \gamma_{2}}{\left[H A_{i}^{-}\right] \gamma_{1}} . \tag{44}
\end{equation*}
$$

At any point of the titration curve, a relatively simple calculation gives

$$
\begin{equation*}
Y=\left(\frac{K_{11}}{K_{21}}\right) C_{1}^{\circ}+C_{2}^{\circ} X, \tag{45}
\end{equation*}
$$

where

$$
\begin{equation*}
Y=\frac{\mathrm{Fa}_{\mathrm{H}}^{2} \gamma_{1} \gamma_{2} \mathrm{D}_{1}}{\mathrm{~b}_{\mathrm{N}} \mathrm{~V}_{0}\left(\mathrm{a}_{\mathrm{H}} \gamma_{2}+2 \mathrm{~K}_{12} \gamma_{1}\right) \mathrm{K}_{21}}, \quad X=\frac{\mathrm{D}_{1}\left(\mathrm{a}_{\mathrm{H}} \gamma_{2}+2 \mathrm{~K}_{22} \gamma_{1}\right)}{\mathrm{D}_{2}\left(\mathrm{a}_{\mathrm{H}} \gamma_{2}+2 \mathrm{~K}_{12} \gamma_{1}\right)} . \tag{46}
\end{equation*}
$$

and

$$
\begin{equation*}
D_{i}=1+\frac{K_{i 1}}{a_{H} \gamma_{1}}+\frac{K_{i 1} K_{i 2}}{a_{H}^{2} \gamma_{2}}, \quad F=b_{N} C_{B} V+\frac{a_{H}}{\gamma_{1}}-\frac{K_{w}}{a_{H} \gamma_{1}} . \tag{47}
\end{equation*}
$$

For dilute solutions ( $\left(<10^{-1} \mathrm{M}\right)$, the activity coefficients $\gamma_{1}$ and $\gamma_{2}$ can be determined from Eq. (6). Concerning the ionic strength I, a simple calculation gives

$$
\begin{align*}
I & =\left[\mathrm{M}^{+}\right]+\left[\mathrm{H}^{+}\right]+\left[\mathrm{A}_{1}^{2-}\right]+\left[\mathrm{A}_{2}^{2-}\right]= \\
& =\mathrm{b}_{\mathrm{N}} \mathrm{C}_{\mathrm{B}} \mathrm{~V}+\frac{\mathrm{a}_{\mathrm{H}}}{\gamma_{1}}+\frac{\mathrm{b}_{\mathrm{N}} \mathrm{~V}_{0}}{\mathrm{a}_{\mathrm{H}}^{2} \gamma_{2}}\left(\frac{\mathrm{~K}_{11} \mathrm{~K}_{12} C_{1}^{\circ}}{\mathrm{D}_{1}}+\frac{\mathrm{K}_{21} \mathrm{~K}_{22} \mathrm{C}_{2}^{\circ}}{\mathrm{D}_{2}}\right) . \tag{48}
\end{align*}
$$

Equation (45) predicts a linear relationship between $Y$ and $X$ with a y-intercept and slope equal to $\left(\mathrm{K}_{11} / \mathrm{K}_{21}\right) \mathrm{C}_{1}^{0}$ and $\mathrm{C}_{2}^{0}$, respectively. Thus, the composition of the mixture can be deduced by linear regression from the coefficients of Eq. (45); the acidity constants $\mathrm{K}_{\mathrm{i} 1}, \mathrm{~K}_{\mathrm{i} 2}$ (with $\mathrm{i}=1,2$ ) being known. This procedure can be realised by the presently proposed iterative method.

Assuming first that $I=b_{N} C_{B} V+a_{H}$, so that $a_{H}=\left[H^{+}\right]$and $\left[A_{1}{ }^{2-}\right],\left[A_{2}{ }^{2-}\right] \approx 0$, it is possible to determine, from the coefficients of Eq. (45) using the experimental V and pH data, approximate values of $\mathrm{C}_{1}^{\circ}$ and $\mathrm{C}_{2}^{\circ}$. Starting from these values and assuming again that $\mathrm{a}_{\mathrm{H}}=\left[\mathrm{H}^{+}\right]$, it is possible to calculate I from Eq. (48) at any chosen point of the titration curve, and hence $\gamma_{1}$ and $\gamma_{2}$ from Eq. (6). These new values of $\gamma_{1}$ and $\gamma_{2}$ can be used to refine values of I and to repeat this until it converges (the criterion being less than $10^{-5}$ difference in I between two subsequent cycles). By repeating these approximations at each point of the titration curve, it is possible to trace the curve $Y=f(X)$ and to determine by least-square method the values of $C_{1}^{\circ}$ and $C_{2}^{\circ}$. These new values can be used to refine the values of $\mathrm{C}_{1}^{0}$ and $\mathrm{C}_{2}^{\circ}$, and to repeat this until they converge (the criterion being less than the $10^{-12}$ difference in $\mathrm{C}_{1}^{\circ}$ and $\mathrm{C}_{2}^{\circ}$ values between two subsequent cycles).

### 2.6. Application of the Iterative Methods for the Analysis of Titration Curves

To establish the reliability of the proposed procedures, first it must be shown that they actually work by comparison with a system where the answer is already known. Experimentally, this implies that the proposed ap-
proaches, applied to previously reported systems, give reasonable (or even better) precision in extraction of the sought parameters. In this respect, it was considered necessary to test all the reported procedures: first, with simulated data corresponding to various values of the titration parameters and, second, with experimental titration data. These tests have revealed that the proposed techniques are fairly applicable ${ }^{23,26}$.
Concerning the application of the iterative methods on simulated $V$ and pH data, it has been found that the proposed techniques are able to extract $\mathrm{V}_{\mathrm{e}}$ and pK values in the case where the simulated data are free of experimental errors ("ideal" data). However, more interesting from the experimental viewpoint is the question of how precisely the proposed methods are able to cope with data containing random extraneous contributions, such as the annoying experimental "noise". To investigate this, a Monte Carlo technique was used, as described in detail elsewhere ${ }^{17}$. The main idea of this procedure has been based on the observation that random experimental errors often closely follow a Gaussian (or normal) distribution. Thus, at each point ( $\mathrm{V}_{\mathrm{i}}$ and $\mathrm{pH}_{\mathrm{i}}$ ) of the "ideal" titration curve $\mathrm{pH}_{\mathrm{i}}=\mathrm{f}\left(\mathrm{V}_{\mathrm{i}}\right)$, a number N of normally distributed random variables $\mathrm{pH}_{\mathrm{ij}}$ with mean $\mathrm{pH}_{\mathrm{i}}$ and standard deviation $S_{i}$ were produced. At the experimental level this implies the realisation of $N$ titrations with a precision equal to $\mathrm{S}_{\mathrm{i}}$. At each volume $\mathrm{V}_{\mathrm{i}}$, the obtained N values $\mathrm{pH}_{\mathrm{ij}}$ were averaged. Evidently, the mean values $\left\langle\mathrm{pH}_{\mathrm{ij}}\right\rangle$ would coincide with the corresponding $\mathrm{pH}_{\mathrm{i}}$ values only in the case where the number N of the produced random variables $\mathrm{pH}_{\mathrm{ij}}$ tends to infinity. Using $\left\langle\mathrm{pH}_{\mathrm{ij}}\right\rangle$ data obtained for $\mathrm{N}=4$, it has been found that the proposed techniques lead with a fair accuracy (error $<1 \%$ ) to $\mathrm{V}_{\mathrm{e}}$ and pK values, even when the simulated pH data are considerably obscured by extraneous noise ( $\mathrm{S}_{\mathrm{i}}=10^{-2} \mathrm{pH}$ unit, namely 5-10 times greater than the accuracy of a precision instrument).

The proposed methods were also applied to various simulated titrations curves, corresponding to different pK values, where the pH data contained a systematic error equal to $10^{-2} \mathrm{pH}$ unit. Experimentally, such errors are usually introduced by an erroneous standardization of the pH meter assembly. A systematic error in the pH values gives rise to relative titration errors $\delta \mathrm{V}_{\mathrm{e}}$ and $\delta \mathrm{pK}$ in the values of $\mathrm{V}_{\mathrm{e}}$ and pK , respectively, determined by $\delta \mathrm{V}_{\mathrm{e}}=$ $\left(\mathrm{V}_{\mathrm{e}}-\mathrm{V}_{\mathrm{e}, \text { syst }}\right) / \mathrm{V}_{\mathrm{e}}$ and $\delta \mathrm{pK}=\left(\mathrm{pK}-\mathrm{pK}_{\text {syst }}\right) / \mathrm{pK}$, where $\mathrm{V}_{\mathrm{e}, \text { syst }}$ and $\mathrm{pK}_{\text {syst }}$ are the values of $\mathrm{V}_{\mathrm{e}}$ and pK determined from data containing a systematic error equal to $10^{-2} \mathrm{pH}$ unit. It has been found that $\delta \mathrm{V}_{\mathrm{e}}$ and $\delta \mathrm{pK}$ depend on the strength of the acid that is being titrated. Also the variation of $\delta \mathrm{V}_{\mathrm{e}}$ as a function of the acid strength presents a maximum in the immediate vicin-
ity of the pK value equal to 2.6 (ref. ${ }^{23}$ ). However, in all cases $\delta V_{\mathrm{e}}$ was less than $0.5 \%$. Hence the proposed techniques are able to extract a $\mathrm{V}_{\mathrm{e}}$ value with only a modest error, even when an important systematic error ( $\delta \mathrm{pH}=$ $10^{-2}$ unit) obscures the pH data. It is worth noting that in the case of diprotic acids with $\mathrm{pK}_{\mathrm{i}}>2.9$, the proposed techniques lead to the exact values of $\mathrm{V}_{\mathrm{e}}$ even when the pH data contain an important systematic error equal to $2 \times 10^{-2} \mathrm{pH}$ unit. For these acids, a rough calibration of the electrode system is sufficient in order to extract the accurate value of the acid concentration. The titration error in the pK value decreases as pK increases. For moderately strong acids (with pK <2.5) this error takes values greater than $1 \%, \delta \mathrm{pK}$ being equal to $10^{-2}$. In contrast, for acids with $\mathrm{pK} \approx 4.8$ (such as the lower members of the series of aliphatic monocarboxylic acids) this error takes values equal to ca $0.2 \%$. For such acids the proposed techniques are able to extract the $\mathrm{V}_{\mathrm{e}}$ and pK values with only a modest error $\left(\delta \mathrm{V}_{\mathrm{e}} \approx\right.$ $0.03 \%$ and $\delta \mathrm{pK} \approx 0.2 \%$ ), even when the pH data contain an important systematic error equal to $10^{-2} \mathrm{pH}$ unit. Finally, for acids with $\mathrm{pK}>3.8$, it can be shown that the titration error in the pK values is approximately equal to the $\delta \mathrm{pH}$ error.

On the other hand, application of the iterative methods on experimental data leads to results that are very close to the corresponding data reported in the literature. As an example, we present here the application of the corresponding procedure on experimental data concerning the titration of a mixture of two weak monoprotic acids. This mixture was prepared by mixing $1.03 \times 10^{-2} \mathrm{M}$ propionic acid ( 30 ml ) with $1.05 \times 10^{-2} \mathrm{M}$ formic acid ( 20 ml ). The mixture was titrated with $1 \times 10^{-1} \mathrm{~m} \mathrm{NaOH}$. As shown in Fig. 1a, the titration curve presents only one pronounced inflection (at V $\approx 5.18$ ml ) corresponding to the total neutralisation of the mixture. The measurements carried out in the acidic range of the mixture are given in Table I.

It is now examined whether it is possible, using these data, to extract the desired $p K$ and concentration values. In a first attempt $R^{2}$ vs $V_{e}$ was varied: the starting value for $\mathrm{C}_{1}^{0}$ being $2 \times 10^{-3} \mathrm{M} ; \mathrm{R}^{2}$ being a measure of the adequacy of the fit of Eq. (29) to the titrated data. The results obtained are presented graphically in Fig. 1b. It should be emphasised that one obtains identical results independently of the starting value for $\mathrm{C}_{1}{ }^{\circ}$.

Figure 1 shows that the graph $\mathrm{R}^{2}=f\left(\mathrm{~V}_{\mathrm{e}}\right)$ presents a pronounced maximum in the vicinity of $\mathrm{V}_{\mathrm{e}}=5.2 \mathrm{ml}$. Further, for $5.08 \leq \mathrm{V}_{\mathrm{e}} \leq 5.28$, it was found that the curve $R^{2}=f\left(V_{e}\right)$ can be perfectly fitted, by the least-square method, to the following polynomial (with the correlation coefficient better than 0.9995):

$$
\begin{align*}
R^{2}=\sum_{n=0}^{6} A_{n} V_{e}^{n}= & -8.8752+5.3268 V_{e}-0.67377 V_{e}^{2}- \\
& -5.1405 \times 10^{-2} V_{e}^{3}+2.8774 \times 10^{-3} V_{e}^{4}+  \tag{49}\\
& +2.9221 \times 10^{-3} V_{e}^{5}-2.8269 \times 10^{-4} V_{e}^{6} .
\end{align*}
$$

Consequently, the exact value of $\mathrm{V}_{\mathrm{e}}$, at which $\mathrm{R}^{2}$ reaches maximum value, can be determined by solving (using any numerical method) Eq. (50).

$$
\begin{equation*}
\frac{d R^{2}}{d V_{e}}=0 \tag{50}
\end{equation*}
$$

The root of this equation was found equal to $\mathrm{V}_{\mathrm{e}}{ }^{\text {max }}=5.172 \mathrm{ml}\left(\mathrm{C}_{\text {tot }}^{0}=1.034 \times\right.$ $10^{-2} \mathrm{~mol} \mathrm{l}^{-1}$ ), which is the best approximation to the desired value of $\mathrm{V}_{\mathrm{e}}$.

In a second step, starting from this value of $\mathrm{V}_{\mathrm{e}}$, the variation of $\hat{\mathrm{C}}_{1}^{\circ}$ with $\mathrm{R}_{\text {cf }}$ was examined. For $6.1 \times 10^{-3} \mathrm{~mol} \mathrm{I}^{-1} \leq \hat{\mathrm{C}}_{1}^{0} \leq 6.25 \times 10^{-3} \mathrm{~mol}^{-1}$, the following polynomial (with the square of the correlation coefficient very close to unity):

$$
\begin{equation*}
10^{3} \hat{C}_{1}^{0}=7.3866-3.7954 R_{c f}+4.9682 R_{c f}^{2}-3.4283 R_{c f}^{3}+1.2084 R_{c f}^{4}-0.17165 R_{\mathrm{cf}}^{5} . \tag{51}
\end{equation*}
$$

Table I
Experimental titration data of an aqueous mixture of $1.029 \times 10^{-2} \mathrm{M}$ propionic acid ( 30 ml ) and $1.051 \times 10^{-2} \mathrm{~m}$ formic acid $(20 \mathrm{ml})$ with $1 \times 10^{-1} \mathrm{~m} \mathrm{NaOH}$ at $\mathrm{T}=298.15 \mathrm{~K}$

| $\mathrm{V}, \mathrm{ml}$ | pH | $\mathrm{V}, \mathrm{ml}$ | pH |
| :---: | :---: | :---: | :---: |
| 1.00 | 3.639 | 2.59 | 4.432 |
| 1.17 | 3.727 | 2.84 | 4.545 |
| 1.31 | 3.800 | 3.04 | 4.638 |
| 1.48 | 3.888 | 3.20 | 4.710 |
| 1.65 | 3.975 | 3.35 | 4.778 |
| 1.80 | 4.050 | 3.52 | 4.860 |
| 1.97 | 4.133 | 3.67 | 4.932 |
| 2.12 | 4.207 | 3.87 | 5.036 |
| 2.27 | 4.278 | 4.01 | 5.107 |
| 2.42 | 4.348 |  |  |

From this polynomial and for $\mathrm{R}_{\mathrm{cf}}=1$ one obtains $\hat{\mathrm{C}}_{1}^{0}=6.168 \times 10^{-3} \mathrm{~mol} \mathrm{l}^{-1}$. This value is the best approximation to the exact value of $\mathrm{C}_{1}^{0}$. Also it results that $\mathrm{C}_{2}^{\circ}=\mathrm{C}_{\text {tot }}^{\circ}-\mathrm{C}_{1}^{0}=4.176 \times 10^{-3} \mathrm{~mol} \mathrm{I}^{-1}$. These values are in excellent agreement with the expected values of $C_{1}^{\circ}\left(=30 \times 0.1029 / 50=6.174 \times 10^{-3}\right.$ $\left.\mathrm{mol} \mathrm{l}^{-1}\right)$ and $\mathrm{C}_{2}^{0}\left(=20 \times 0.01051 / 50=4.204 \times 10^{-3} \mathrm{~mol}^{-1}\right)$. Finally, using the determined values of $\mathrm{V}_{\mathrm{e}}$ and $\mathrm{C}_{1}^{0}$, one obtains from the coefficients of Eq. (23) the values $\mathrm{pK}_{1}=4.877$ and $\mathrm{pK}_{2}=3.742$. These values are very close to the corresponding values ( 4.874 and 3.752 , at $25^{\circ} \mathrm{C}$ ) reported in the literature ${ }^{20,27}$.


Fig. 1
Titration curve of an aqueous mixture of $1.029 \times 10^{-2} \mathrm{M}$ propionic acid ( 30 ml ) and $1.051 \times$ $10^{-2} \mathrm{M}$ formic acid ( 20 ml ) with $1 \times 10^{-1} \mathrm{M} \mathrm{NaOH}(\mathrm{a})$, and variation of $\mathrm{R}^{2}$ with $\mathrm{V}_{\mathrm{e}}(\mathrm{b})$

## 3. DETERMINATION OF KINETIC PARAMETERS FROM CHRONOAMPEROMETRIC DATA

### 3.1. Theoretical Background

Potential step chronoamperometry has proved to be a valuable technique for the study of various electrochemical reactions because of the relative simplicity of the mathematics involved in solving the appropriate diffusion equations. Some typical examples, for which the application of the above-mentioned technique leads to analytical solutions relating the current i with time t , are the reaction schemes (I)-(V).

$$
\begin{align*}
& A \stackrel{n_{1} e}{\Longleftrightarrow} B \xrightarrow{k_{1}} C \stackrel{n_{2} e}{\Longleftrightarrow} D \xrightarrow{k_{2}} C \stackrel{n_{3} e}{\Longleftrightarrow} F  \tag{I}\\
& A \stackrel{n_{1} e}{\Longleftrightarrow} B \xrightarrow{k_{1}} C \stackrel{n_{2} e}{\Longleftrightarrow} D  \tag{II}\\
& \mathrm{O}+\mathrm{ne} \xlongequal{\mathrm{k}_{\mathrm{f}}} \mathrm{R}  \tag{III}\\
& O+n e \underset{k_{b}}{\stackrel{k_{f}}{\Longrightarrow}} R  \tag{IV}\\
& \mathrm{O} \xrightarrow[\mathrm{k}_{-1}]{\stackrel{\mathrm{k}_{1}}{ }} \mathrm{P} \underset{\text { fast }}{\text { ne }} \mathrm{R} \tag{V}
\end{align*}
$$

The theoretical treatment of the so-called ECECE mechanism (reaction scheme (I)) for potential step chronoamperometry at a planar electrode, the initial and boundary value problems, treated by means of the Laplace transform method, led to the following relation ${ }^{16}$ :

$$
\begin{equation*}
\frac{\mathrm{i}}{\mathrm{n}_{1} \mathrm{FAD}^{1 / 2} \mathrm{c}_{A}^{\mathrm{b}}}=\left(1+\frac{\mathrm{n}_{2}}{\mathrm{n}_{1}} \Phi_{1}(\mathrm{t})+\frac{\mathrm{n}_{3}}{\mathrm{n}_{1}} \Phi_{2}(\mathrm{t})\right) \frac{1}{\sqrt{\pi \mathrm{t}}}, \tag{52}
\end{equation*}
$$

where $c_{A}^{b}$ is the bulk concentration of species $A, F$ is Faraday constant, $A$ is the electrode area, and $D$ is the assumed common diffusion coefficient of all the species involved in the ECECE sequence. The functions $\Phi_{1}(\mathrm{t})$ and $\Phi_{2}(\mathrm{t})$ are defined by the relations:

$$
\begin{gather*}
\Phi_{1}(\mathrm{t})=1-\mathrm{e}^{-k_{1} \mathrm{t}}  \tag{53}\\
\Phi_{2}(\mathrm{t})=1-\mathrm{e}^{-\mathrm{k}_{1} \mathrm{t}}-\frac{\sqrt{\mathrm{t}}}{2} \exp \left(-\frac{\mathrm{k}_{1}+\mathrm{k}_{2}}{2} \mathrm{t}\right) \mathrm{X}(\mathrm{t}),
\end{gather*}
$$

where

$$
\begin{equation*}
X(t)=\int_{0}^{t} \frac{\left(1-e^{-k_{1} y}\right) \exp \left(\frac{k_{1}+k_{2}}{2} y\right)}{y^{3 / 2}} I_{0}\left(\frac{k_{1}-k_{2}}{2}(t-y)\right) d y \tag{54}
\end{equation*}
$$

and $I_{0}(x)$ is the modified Bessel function of zero order, $x$ being the argument $\left\{\left(\mathrm{k}_{1}-\mathrm{k}_{2}\right) / 2\right\}(\mathrm{t}-\mathrm{y})$.

For the ECE mechanism (reaction scheme (II), the current-time relation is given by (ref. ${ }^{14}$ ):

$$
\begin{equation*}
\frac{i}{n_{1} F A D^{1 / 2} C_{A}^{b}}=\left(1+\frac{n_{2}}{n_{1}}\left(1-e^{-k t}\right)\right) \frac{1}{\sqrt{\pi t}} . \tag{55}
\end{equation*}
$$

The chronoamperometric relations for the reaction schemes (III) (irreversible mechanism), (IV) (quasi-reversible mechanism) and (V) (CE mechanism) have the respective forms ${ }^{14}$

$$
\begin{gather*}
i=n F A c_{0}^{b} k_{f} \exp \left(\beta_{1}^{2} t\right) \operatorname{erfc}\left(\beta_{1} \sqrt{t}\right)  \tag{56}\\
i=n F A c_{0}^{b} k_{f} \exp \left(\beta_{2}^{2} t\right) \operatorname{erfc}\left(\beta_{2} \sqrt{t}\right)  \tag{57}\\
i=n F \beta_{3} c_{0}^{b} A \sqrt{D} \exp \left(\beta_{3}^{2} t\right) \operatorname{erfc}\left(\beta_{3} \sqrt{t}\right), \tag{58}
\end{gather*}
$$

where $\beta_{1}=k_{f} / \sqrt{D}, \beta_{2}=\left(k_{f}+k_{b}\right) / \sqrt{D}, \beta_{3}=k_{1} / \sqrt{k_{-1}}$, provided that $k_{-1} \gg k_{1}$, and $D$ is the assumed common diffusion coefficient of all the species involved in reaction schemes (III)-(V).

The reaction schemes (III)-(V) share a chronoamperometric response of the form

$$
\begin{equation*}
\mathrm{i}=\mathrm{i}_{0} \exp \left(\beta^{2} \mathrm{t}\right) \operatorname{erfc}(\beta \sqrt{\mathrm{t}}), \tag{59}
\end{equation*}
$$

where $i_{o}$ is the initial current, usually not accessible by direct measurement.

Equation (52) can be applied for studying the kinetics of homogeneous chemical reactions involved in the ECECE mechanism. However, because of its non-linear form, the determination of the rate constants from $i=f(t)$ curves using classic numerical methods appears to be very difficult. That was the reason why in a previous investigation ${ }^{16}$ we chose alternative methods for the determination of the rate constants of the intervening chemical reactions in the ECECE mechanism of 1,4-benzoquinone dioxime ( $p-B Q D$ ). Nevertheless, it is not always possible to find such alternative methods for determining the rate constants in any ECECE mechanism.

Equation (59) also has a non-linear form. Thus, the determination of the corresponding rate constants from $i=f(t)$ plots by using traditional numerical methods without prior knowledge of $A, D$ or $i_{0}$ appears to be very difficult. For this reason, various approximate methods for the determination of the rate constants have been suggested in the literature ${ }^{44-48}$. These methods, using either disc or ring-disc measurements, or chronoamperometric data at short or long times, allow to evaluate the kinetic parameters with only low precision. Of course, there exist numerical methods that can extract from Eq. (59) the accurate values of the corresponding kinetic parameter ${ }^{14,49}$. However, these methods (based on the linearisation of the chronoamperometric data either in Laplace space or by calculating the semiintegral of the current) are of a complexity that does not appeal to the average chemist. An alternative technique to these methods has been devised in the literature ${ }^{50}$. However, this procedure is of experimental complexity, since it requires combination of chronoamperometric and chronocoulometric data.

This situation has encouraged us to develop new iterative methods, based on Eqs (52), (55), and (59) that allow determination of the exact values for the corresponding rate constants exclusively from chronoamperometric data ${ }^{15-17}$.

### 3.2. Determination of the Rate Constants $\mathrm{k}_{1}$ and $\mathrm{k}_{2}$ of an ECECE M echanism

This method is based on the main principles of a previous one ${ }^{31,51}$ that was used for the determination of the rate constants for two competitive parallel reactions preceded by an equilibrium between two of the reactants.

Equation (52) can be rearranged to give the following form:

$$
\begin{equation*}
Y=b X, \tag{60}
\end{equation*}
$$

where

$$
\begin{equation*}
Y=\frac{i \sqrt{t}}{n_{1} F A D^{1 / 2} c_{A}^{b}}, \quad X=1+\frac{n_{2}}{n_{1}} \Phi_{1}(t)+\frac{n_{3}}{n_{1}} \Phi_{2}(t) . \tag{61}
\end{equation*}
$$

Equation (60) predicts a linear relationship between $Y$ and $X$ with a gradient equal to $1 / \sqrt{\pi}$. Alternatively, the experimental confirmation of such a correlation supports the assumption that the reduction under consideration does, in fact, follow an ECECE mechanism.

However, despite the fact that plots of $Y$ vs $X$ offer a theoretical basis for the correlation of the results, their practical application may be regarded as fairly limited. Indeed, calculation of $X$ at various times requires estimates of $k_{1}$ and $k_{2}$ that are generally not available.

However, as mentioned above, approximate values of these constants could be obtained in certain instances by alternative methods. We examine now whether it is possible, starting from these values, to determine the exact $k_{1}$ and $k_{2}$ values, using only experimental chronoamperometric data.
We assume first that the values of $k_{1}$ and $k_{2}$, a priori accessible, are defined by a set S including an infinity of elements. Each element of S corresponds to a pair of values of $k_{1}$ and $k_{2}$. Consequently, the various elements of $S$ define various electrochemical reactions following the ECECE mechanism. Using an element of S (namely a pair of $\mathrm{k}_{1}$ and $\mathrm{k}_{2}$ values chosen arbitrarily), it is possible to trace, using the chronoamperometric data, the curve $\mathrm{Y}=$ $f(X)$. This will approach a straight line to the extent where the chosen values of the rate constants also approach the exact $k_{1}$ and $k_{2}$ values. Thus, the best linearity that could be obtained, using the available experimental data, corresponds evidently to the best approximation to the exact values of the sought rate constants.

However, among infinity of elements of S, there is of course one corresponding to the electrochemical reaction under consideration. In an attempt to determine the values of the rate constants corresponding to this element, we assume that we have an initial estimation of either $k_{1}$ or $k_{2}$. Let $\mathrm{k}_{1}^{\circ}$ be an approximate value of $\mathrm{k}_{1}$. This value can be obtained by an alternative method. Starting from this value, it is possible to trace N curves $\mathrm{Y}=$ $f(X), N$ being the number of $k_{2}$ values taken for these calculations. For each of these curves, the calculation of the squared correlation coefficient $R^{2}$ allows to compare the linearity of the various $Y=f(X)$ plots and to trace the curve $R^{2}=f\left(k_{2}\right)$. This curve is expected to reach a pronounced maximum at a value of $k_{2}$ equal to $k_{2}^{1}$, which can be considered as a first approximation to the exact $\mathrm{k}_{2}$ value.

In a second step, starting from $k_{2}^{1}$, it is possible to trace the curve $R^{2}=$ $f\left(k_{1}\right)$ and to determine, in the same way, the first approximation $k_{1}^{1}$ to the $k_{1}$ value.
This procedure can be repeated $n$ times in order to obtain the sequences $\left\{k_{1}^{n}\right\}$ and $\left\{k_{2}^{n}\right\}$ that converge to the limiting values $k_{1}^{\text {lim }}$ and $k_{2}^{\text {lim }}$, expressing the best approximation to the exact values of the corresponding rate constants. It should be noted that as $n$ grows, $k_{2}^{1}$ and $k_{2}^{n}$ are expected to be increasingly better estimates of the desired rate constants. One stops the iterations when sufficient accuracy is obtained.

### 3.3. Determination of the Rate Constants of ECE, Irreversible, Quasi-Reversible, and CE Mechanisms

Equations (55) and (59) can be written as

$$
\begin{equation*}
y=B x, \tag{62}
\end{equation*}
$$

where (for Eq. (55))

$$
\begin{equation*}
y=\frac{i \sqrt{\pi t}}{n_{1} F c_{A}^{b}}, \quad x=1+\frac{n_{2}}{n_{1}}\left(1-e^{-k t}\right), \quad B=A \sqrt{D} \tag{63}
\end{equation*}
$$

(for Eq. (59))

$$
\begin{equation*}
y=i, \quad x=\exp \left(\beta^{2} t\right) \operatorname{erfc}(\beta \sqrt{t}), \quad B=i_{0} . \tag{64}
\end{equation*}
$$

Equation (62) predicts a linear relationship between $y$ and $x$. As in the case of the ECECE mechanism, the experimental confirmation of such a correlation, using the above equations, supports the assumption that the reaction under consideration follows, indeed, the corresponding mechanism. However, there is no way in which, without prior knowledge of the kinetic parameters, the $y$ vs $x$ plots give a linear graph. This is possible with the proposed iterative method.

We assume first that the value of the corresponding kinetic parameter $k^{\prime}$ ( $k^{\prime}$ denoting $k$ or $\beta$ ) lies in an interval ( $a, b$ ). Choosing arbitrarily from this interval a value for $k^{\prime}$, it is possible to trace, using the chronoamperometric data, the curve $y=f(x)$. This will approach a straight line to the extent where the chosen value for $k^{\prime}$ also approaches the exact value of the corresponding kinetic parameter. Thus, the best linearity that could be obtained,
using the available experimental data, corresponds evidently to the best approximation to the exact value of $k^{\prime}$. Hence, by seeking within the interval $(a, b)$ the $k^{\prime}$ value, it is possible to trace $N$ curves $y=f(x), N$ being the number of $k^{\prime}$ values taken for these calculations. For each of these curves, the calculation of the squared correlation coefficient $R^{2}$ allows to compare the linearity of various $y=f(x)$ plots and trace the curve $R^{2}=f\left(k^{\prime}\right)$. This curve is expected to present a pronounced maximum at a value of $\mathrm{k}^{\prime}$ equal to $\mathrm{k}_{\max }^{\prime}$, which can be considered as the best approximation to the exact value of the kinetic parameter. The slope of the straight line corresponding to $\mathrm{k}_{\max }^{\prime}$ is the best approximation to the exact value of $B$ (being equal to $A \sqrt{D}$ or $i_{0}$ ).

### 3.4. Application of the Iterative $M$ ethods for Determination of Kinetic Parameters from Chronoamperometric Data

All the methods described herein were applied to simulated chronoamperometric data, either without or with extraneous "noise" (M onte Carlo simulated data), for different values of the corresponding kinetic parameters ${ }^{15,17}$. The Monte Carlo data were produced as described previously in the case of the analysis of titration data. The obtained results showed that these procedures are able to extract the kinetic parameters with only modest errors, even when the faradaic data are considerably obscured by random extraneous contributions, such as annoying experimental "noise". The reported techniques were also successfully applied to experimental data ${ }^{15,17}$. As an example, we present here the determination of $k_{1}$ and $k_{2}$ of an ECECE mechanism, using experimental data concerning the reduction of 1,2-benzoquinone dioxime ( $10^{-3} \mathrm{M}$ ) in aqueous $5 \times 10^{-1} \mathrm{M} \mathrm{HClO} 4$ and $2.5 \times$ $10^{-4} \mathrm{M} \mathrm{Bi}\left(\mathrm{ClO}_{4}\right)_{3}$ solution at a Pt electrode modified by underpotential deposition (upd) of Bi at $25^{\circ} \mathrm{C}$. The reduction of 1,2-benzoquinone dioxime (o-BQD) at a $\mathrm{Pt} / \mathrm{Bi}(\mathrm{upd})$ modified electrodes in acidic solutions proceeds via the mechanism shown in Scheme $1^{18}$.



[^0]Mean values of i obtained from three independent experiments are given in Table II. The values of Y calculated from Eq. (61) are also listed in Table II. In these calculations, the effective (real) electrode area A and the diffusion coefficient D were taken equal to $0.136 \mathrm{~cm}^{2}$ and $6.80 \times 10^{-6} \mathrm{~cm}^{2} \mathrm{~s}^{-1}$, respectively ${ }^{52}$. Using the values of $Y$ reported in Table II, the following smoothing function was found by the least-square method ( $R^{2}>0.9993$ ):

$$
Y=0.65249+10.786 t-69.413 t^{2}+259.55 t^{3}-497.54 t^{4}+375.53 t^{5} .(65)
$$

Smoothed $Y$ values ( $Y_{\text {sm }}$ ), calculated from Eq. (65), were used in the application of the iterative procedure for the determination of the rate constants $k_{1}$ and $k_{2}$. It is noted that the $Y_{s m}$ values, used in the iterations, ranged from

Table II
Experimental current values for the reduction of o-BQD at a $\mathrm{Pt} / \mathrm{Bi}$ (upd) electrode in aqueous acid solutions and the values of $Y$ calculated from Eq. (61) at $T=298.15 \mathrm{~K}$

| $\mathrm{t}, \mathrm{s}$ | $\mathrm{i}, \mu \mathrm{A}$ | Y |
| :---: | :---: | :---: |
| 0.05 | 320.39 | 1.04615 |
| 0.08 | 287.72 | 1.18834 |
| 0.10 | 271.05 | 1.25164 |
| 0.13 | 251.01 | 1.32157 |
| 0.15 | 239.62 | 1.35517 |
| 0.18 | 227.51 | 1.40952 |
| 0.20 | 220.27 | 1.43845 |
| 0.23 | 209.48 | 1.46704 |
| 0.25 | 204.47 | 1.49289 |
| 0.28 | 195.52 | 1.51079 |
| 0.30 | 191.70 | 1.53322 |
| 0.33 | 184.48 | 1.54751 |
| 0.35 | 180.63 | 1.56042 |
| 0.38 | 174.77 | 1.57322 |
| 0.40 | 170.99 | 1.57917 |

0.05 to 0.35 s , since in this region the process under consideration shows a pronounced kinetic character.
The rate constants of the coupled chemical reactions could be determined exclusively from chronoamperometric data (the iterative method reaches the exact values of $k_{1}$ and $k_{2}$ even if one starts from very poor estimates of either $\mathrm{k}_{1}$ or $\mathrm{k}_{2}$ ). However, approximate values at least for one rate constant should be useful, since the number of iterations required is smaller when the starting value is closer to its real magnitude. For this reason, the approximate value $\mathrm{k}_{1}=26.14 \mathrm{~s}^{-1}$, determined previously ${ }^{18}$, was used as the starting value of the iterative technique. The $\left\{k_{1}^{n}\right\}$ and $\left\{k_{2}^{n}\right\}$ sequences are summarised in Table III. It should be noted that in all cases the variations $R^{2}=f\left(k_{j}\right)$ (with $j=1$ or 2 ) showed pronounced maxima, such as plotted in Fig. 2a.
As can be deduced from Table III, one practically reaches the desired values after six iterations. The mean value obtained (for $n>6$ ) was equal to $k_{1}^{n}$ $=28.25 \pm 0.01 \mathrm{~s}^{-1}$ and $\mathrm{k}_{2}^{\mathrm{n}}=7.488 \pm 0.003 \mathrm{~s}^{-1}$. The small fluctuations observed around the limiting values for $n>6$ can be attributed to round-off

## Table III

Successive approximations to $\mathrm{k}_{1}$ and $\mathrm{k}_{2}$ values obtained by the iterative method for an ECECE mechanism and using the experimental data reported in Table II. T = 298.15 K

| $n$ | $k_{1}^{n}, s^{-1}$ | $k_{2}^{n}, s^{-1}$ |
| :---: | :--- | :--- |
| 0 | 26.14 |  |
| 1 | 26.738 | 6.185 |
| 2 | 27.411 | 6.621 |
| 3 | 27.892 | 7.065 |
| 4 | 28.126 | 7.331 |
| 5 | 28.210 | 7.437 |
| 6 | 28.239 | 7.474 |
| 7 | 28.249 | 7.484 |
| 8 | 28.247 | 7.489 |
| 10 | 28.254 | 7.487 |
| 11 | 28.258 | 7.490 |

errors, introduced mainly in the determination of the maxima of the $R^{2}=$ $\mathrm{f}\left(\mathrm{k}_{\mathrm{j}}\right)$ curves.

To test the exactness of the values derived using Eq. (52), the curve $i=f(t)$ was traced and compared with experimental $i$ values. The agreement is excellent (Fig. 2b).

On the other hand, the determination of $X$ as a function of $t$, using Eq. (61) and the values for $k_{1}$ and $k_{2}$ extracted above, allows to correlate the variation of $Y$ with $X$. This variation was found to be perfectly linear, the



Fig. 2
Chronoamperometric response for the reduction of o-BQD at a $\mathrm{Pt} / \mathrm{Bi}$ (upd) electrode in aqueous acid solution. Variation of $\mathrm{R}^{2}$ with $\mathrm{k}_{2}\left(\mathrm{k}_{1}=\mathrm{k}_{1}^{\circ}=26.14 \mathrm{~s}^{-1}\right)(\mathrm{a})$, and variation of the current with time (b). Theoretical curve calculated using $\mathrm{k}_{1}=28.25 \mathrm{~s}^{-1}$ and $\mathrm{k}_{2}=7.488 \mathrm{~s}^{-1}(-)$, experimental values (■)
corresponding correlation coefficient being very close to unity. The slope b of this dependence, found equal to 0.5611 , is very close to the theoretical one ( $b=1 / \sqrt{\pi}=0.5642$ ). This behaviour provides further experimental evidence that the reduction of o-BQD at a $\mathrm{Pt} / \mathrm{Bi}(u p d)$ electrode proceeds via an ECECE mechanism.

## 4. CONCLUSIONS

Theory and iterative methods for analysis of potentiometric titration data are presented in this paper. The proposed methods, using equations derived without approximations, and data exclusively resulting from the buffer region of a titration curve, enable the prediction of the accurate values of the concentration(s) and the thermodynamic dissociation constant(s) of the titrated acid(s), even if the titration is stopped well before the end-point of the titration. All these methods can successfully be applied in titrations without any sharp end-point, or for the determination of the analytical concentration and the thermodynamic pK value for a moderately weak acid in the presence of other very weak acids, the neutralisation of which is undesirable. The last determination can easily be done by stopping the titration well before the end-point. The majority of the methods reported in the literature are valid when (i) the titrations are carried out under conditions of constant ionic strength, and (ii) the accurate value of the analytical concentration of the titrated acid is known. On the contrary, the proposed procedures described above are valid even when (i) the titrations are performed under conditions of varying ionic strength (but, there is no reason why the analysis would not be carried out under conditions of constant ionic strength), and (ii) the accurate value of the analytical concentration of the titrated acid is not known. Hence, the addition of a neutral salt to the titrated solution to keep the ionic strength constant, is not necessary. Therefore, the pK values obtained in media of a low ionic strength are free of any possible "salt effect". Another advantage of the proposed techniques, in comparison to other approaches in the literature, is the simultaneous determination of the accurate values of the analytical concentration and the corresponding thermodynamic pK value of the titrated acid. Indeed, thermodynamic pK values being independent of the concentration could be considered as the "fingerprint" of the titrated acid. Thus, from the comparison of the determined pK values with the tabulated ones it is possible to identify the titrated acid.

Iterative methods for evaluating the chronoamperometric curves of various electrochemical mechanisms are proposed. Thus, in the case of an

ECECE mechanism, the proposed method allows to determine the exact values of the corresponding rate constants $k_{1}$ and $k_{2}$. The test of this procedure, using simulated ideal data (free of any extraneous "noise"), showed that this method is fairly well applicable. Application of the iterative technique to Monte Carlo simulated data revealed that the proposed procedure is also able to extract kinetic parameters even if the faradaic data are considerably obscured by "noise". Finally, this method was successfully applied to experimental chronoamperometric data.
The proposed method for the analysis of chronoamperometric data of various electrochemical mechanisms (ECE, irreversible, quasi-reversible, and CE mechanisms) is also applicable even if the faradaic data are considerably obscured by gaussian "noise". The main advantage of this iterative method, in comparison with other methods currently in use, is that the accurate values of the rate constants of the coupled chemical reactions in the CE and ECE mechanisms can be obtained directly from the i-t data even if the parameters A (the effective electrode area) and D are not known. These parameters are usually not available.

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[^0]:    Scheme 1

